

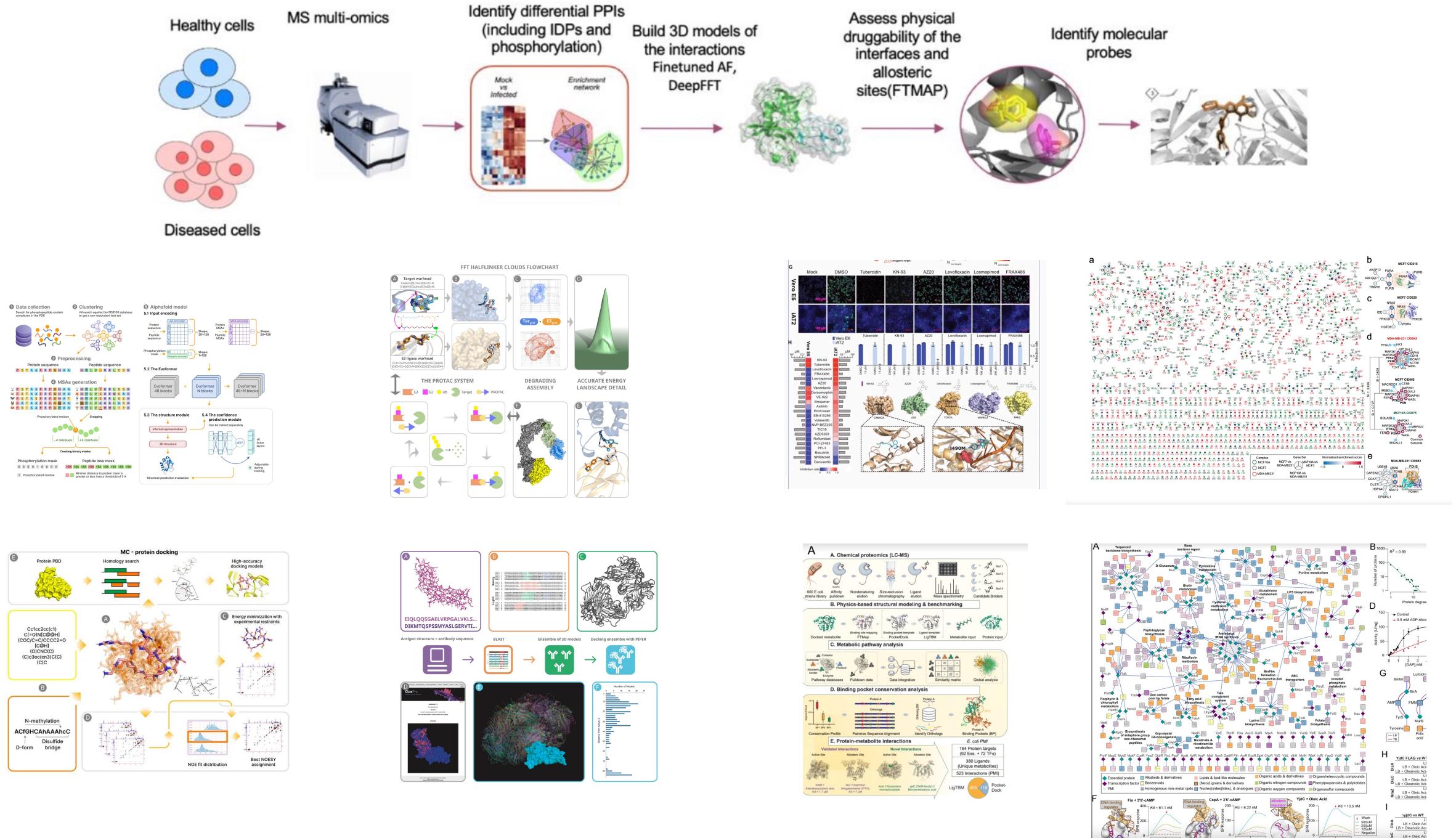
# **Systems level understanding and regulating of the disease with atomic resolution**



**Stony Brook  
University**

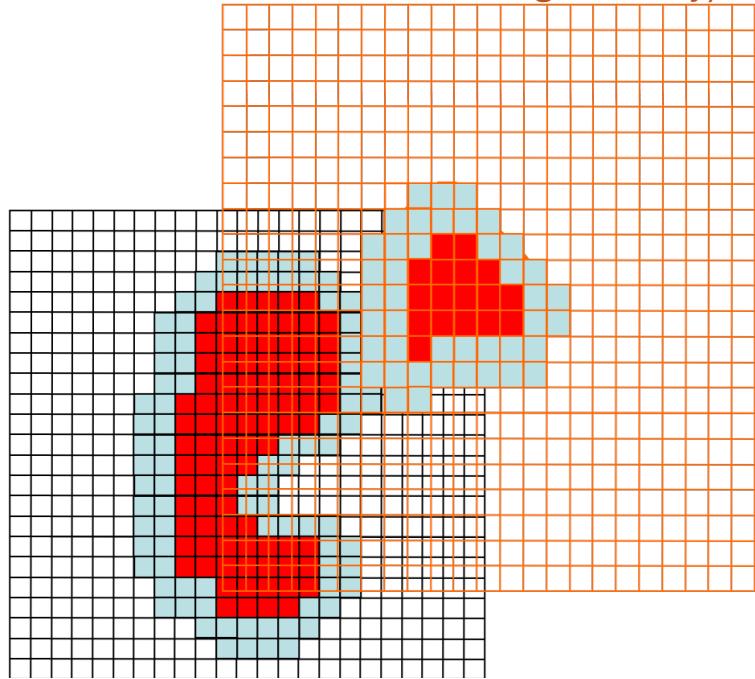


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FOR PHYSICAL AND QUANTITATIVE BIOLOGY



# FFT convolution approach enables global exhaustive macromolecular interaction sampling

Ligand representation (e.g.  
charge density)



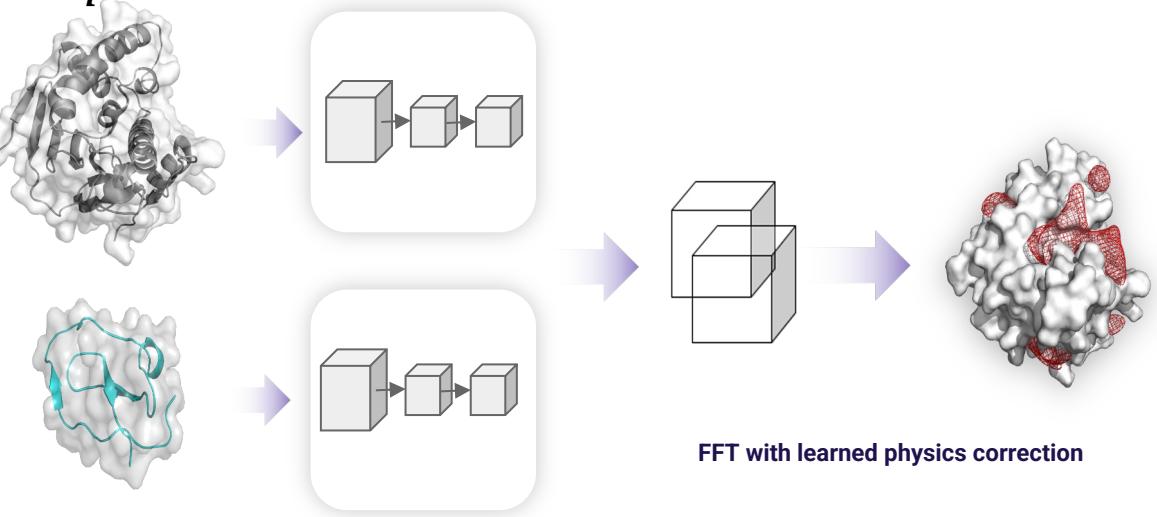
Receptor representation  
(e.g. electrostatic  
potential)  $O(N^6) \rightarrow O(N^3 \ln N^3)$

Interaction energy as a sum of FFT convolutions

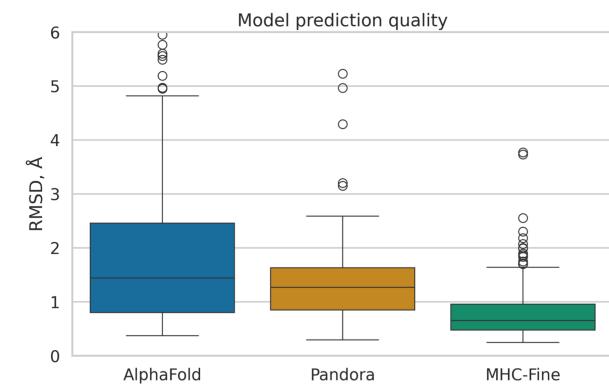
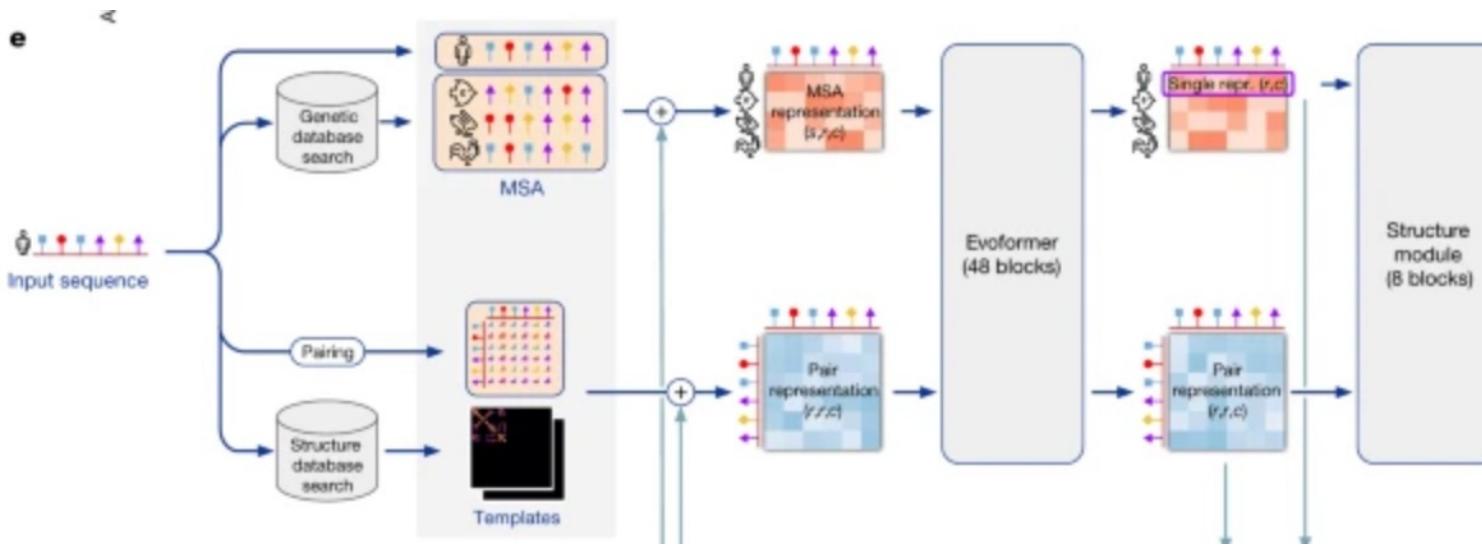
$$E(\bar{t}, r) = \sum_n \sum_{\bar{x}} R_p(\bar{x}) L_{pr}(\bar{t} - \bar{x})$$

$$E(\bar{t}, r) = IFT \left[ \sum_p FT^* \{R_p(\bar{x})\} FT \{L_{pr}(\bar{x})\} \right]$$

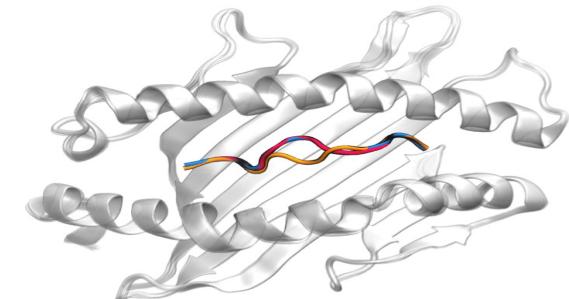
$$\begin{aligned} E_{full} &= E_{vdw} + E_{elec} + E_{pair} \\ E_{vdw} &= E_{rep} + E_{attr} \\ E_{elec} &= \sum_{i=1}^{N_1} \sum_{j=1}^{N_2} \frac{q_i q_j}{(r_{ij}^2 + D^2 \exp(-\frac{r_{ij}^2}{4D^2}))^{1/2}} \\ E_{pair} &= \sum_{i=1}^{N_1} \sum_{j=1}^{N_2} \varepsilon(i, j) = \begin{cases} 0, r_{ij} > D \\ \varepsilon_{ij}, d < r_{ij} < D \\ \varepsilon_{ij} = \sum_t \varepsilon_{ti} \lambda_t \varepsilon_{tj} \end{cases} \end{aligned}$$



# Pytorch-AF – Customized Alphafold-style architecture

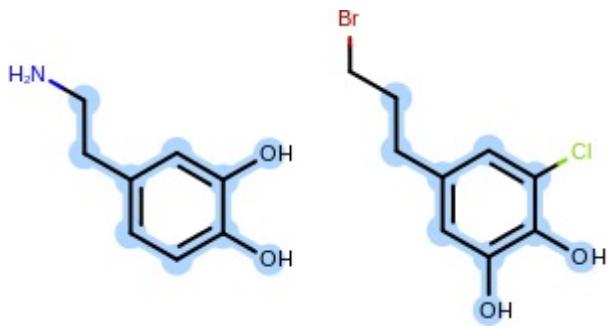


Jumper et. al. 2021; Glukhov et. al 2023

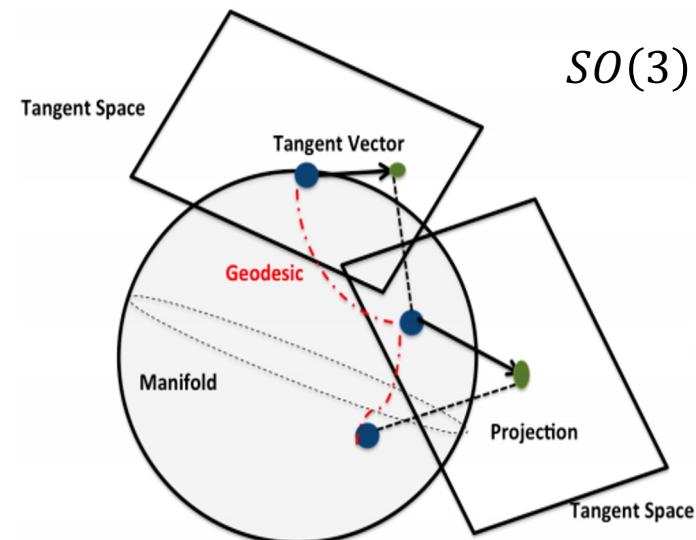


# LigTBM protein-ligand docking

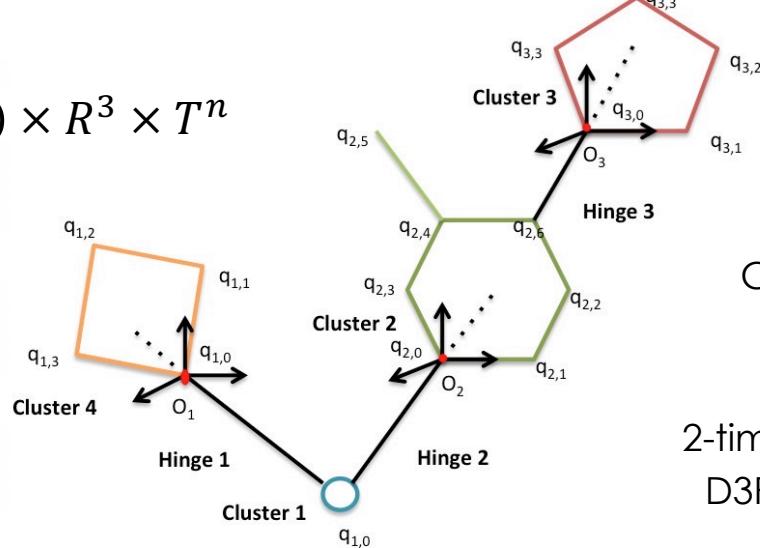
## Maximum Common Substructure



## Diffusion on the manifolds

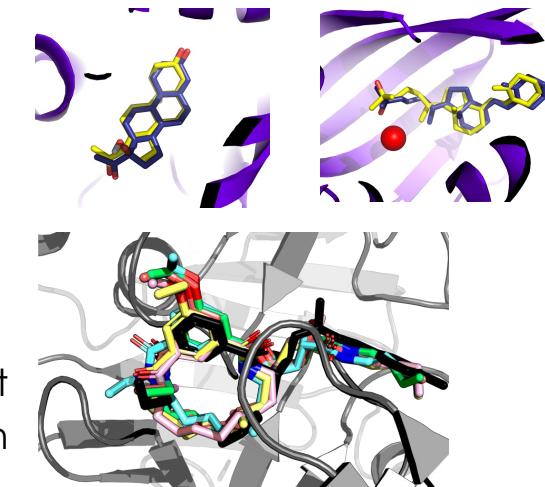
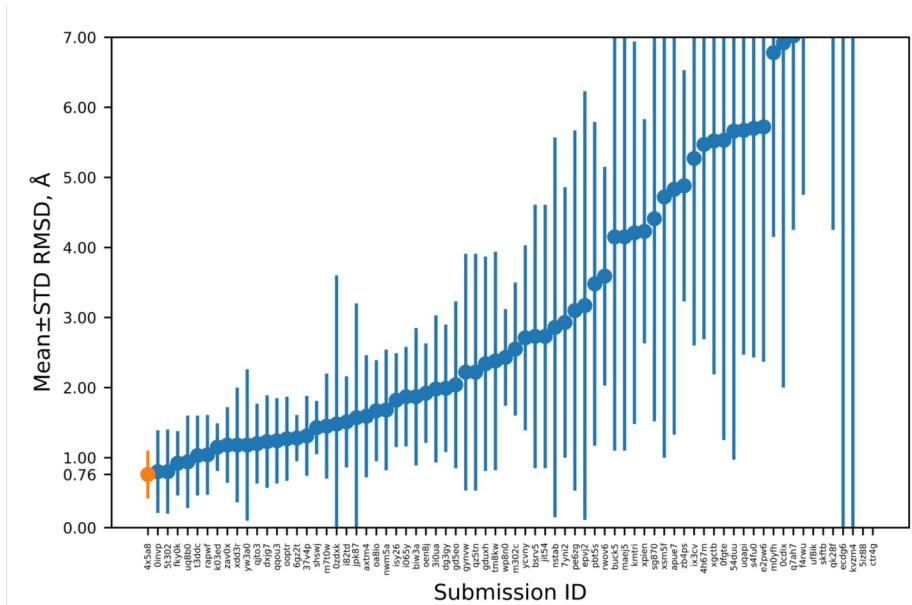


$$SO(3) \times R^3 \times T^n$$



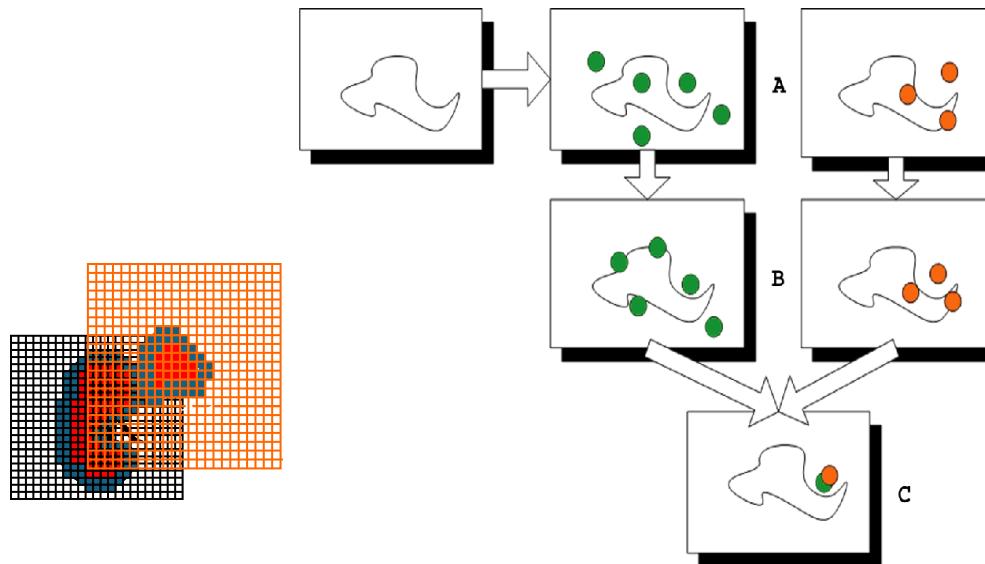
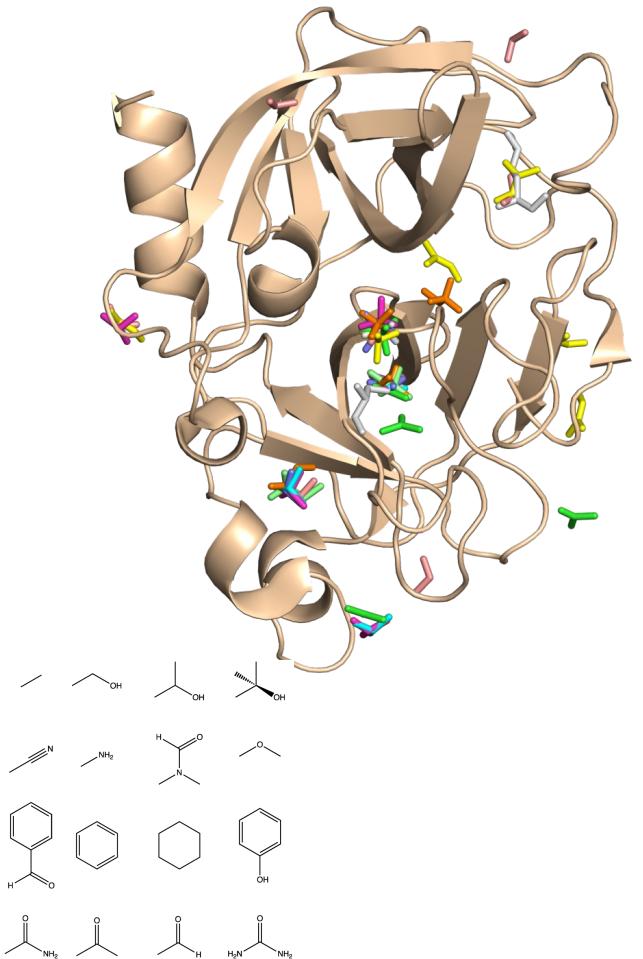
Top performer in the latest  
CASP (ligand prediction) and  
GPCR Dock competitions

2-times top performer in the latest  
D3R ligand docking competition



Padhorny et al., JCAMD 2018; Ignatov et al., JCAMD 2018; Kotelnikov et al., JCAMD 2019;  
Alekseenko et al., JMB 2020; Kotelnikov et al., Proteins 2023;

# FTMap – computational solvent mapping

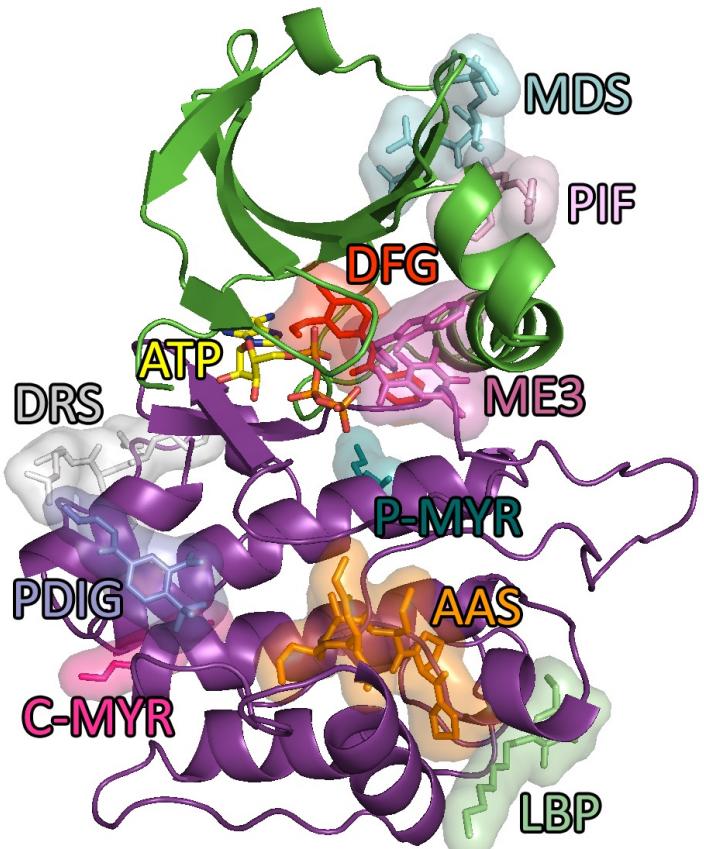


Druggable sites bind a variety of small molecules

“Hit rate” is a predictor of druggability

Brenke et. al 2009 Bioinformatics; Kozakov et al PNAS 2011;  
Villar et al Nature Chem Bio 2014; Kozakov et al. Nature Protocols 2015;  
Kozakov et. al PNAS 2015; Kozakov et. al J Med Chem 2015; Beglov et. al PNAS 2018; Yueh. J Med. Chem 2019; Egbert et. al 2021; Khan et. al 2023

# Kinase Allostery Atlas



PDB Contains more than 3000 structures

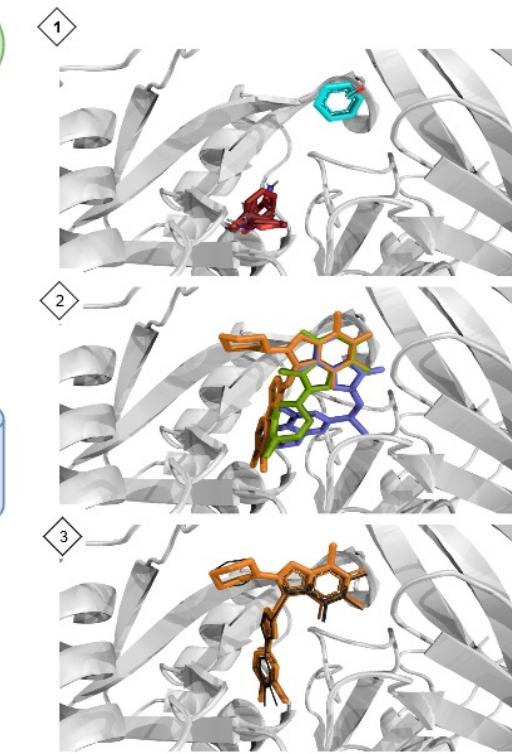
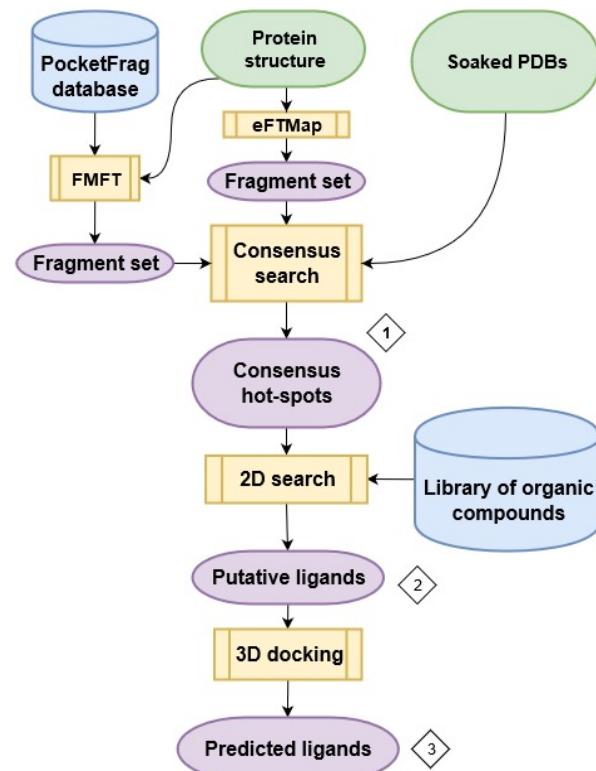
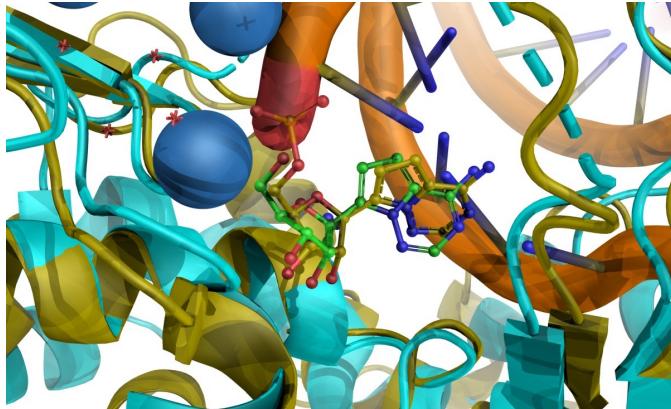
325 Different kinase families

~250 human kinases + 250 more AF structures

Known regulatory sites:

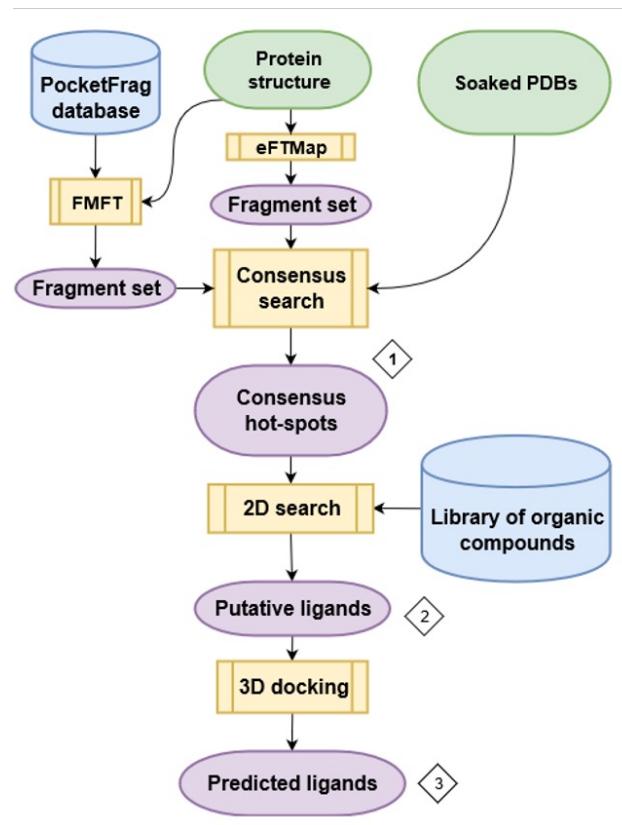
- 1) DFG loop pocket
- 2) PIF pocket few compounds reported for PDK1
- 3) Several others, which are not validated

# Screening of Giga-size libraries



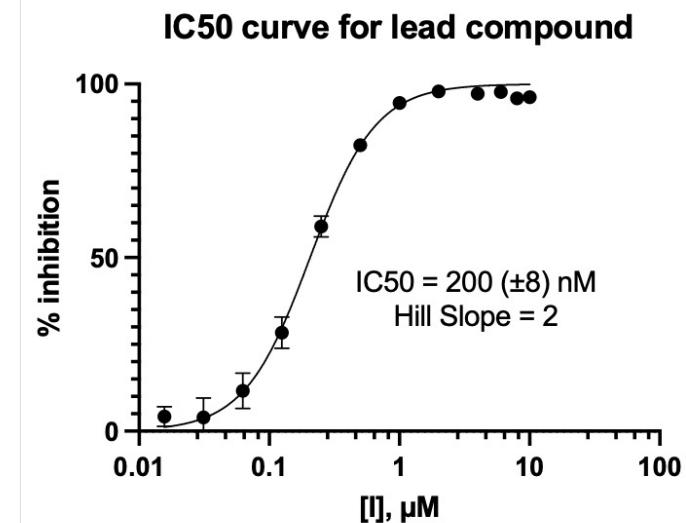
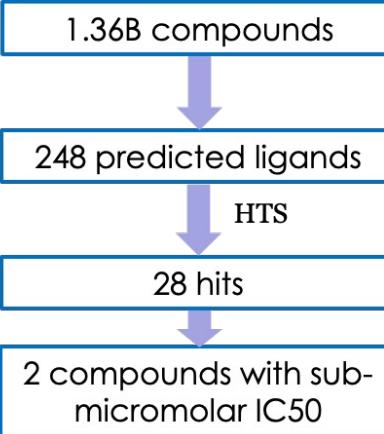
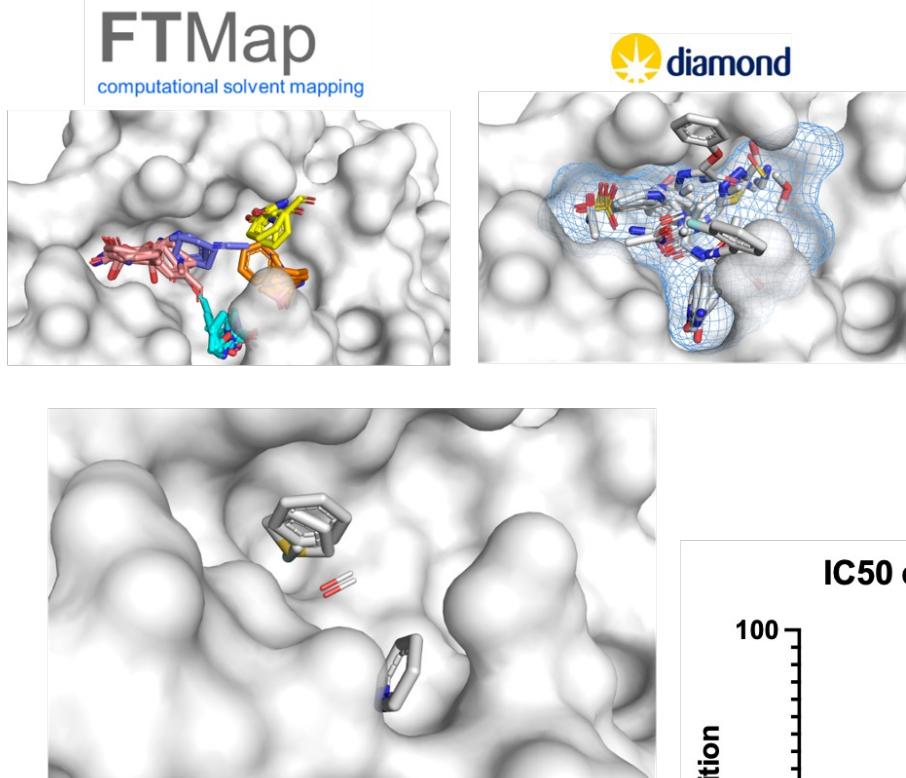
Identified nanomolar and low macromolar hits to a number of COVID targets NSP3, NSP13, Mpro

# Virtual screening of giga-size libraries – SARS-CoV-2 Mpro

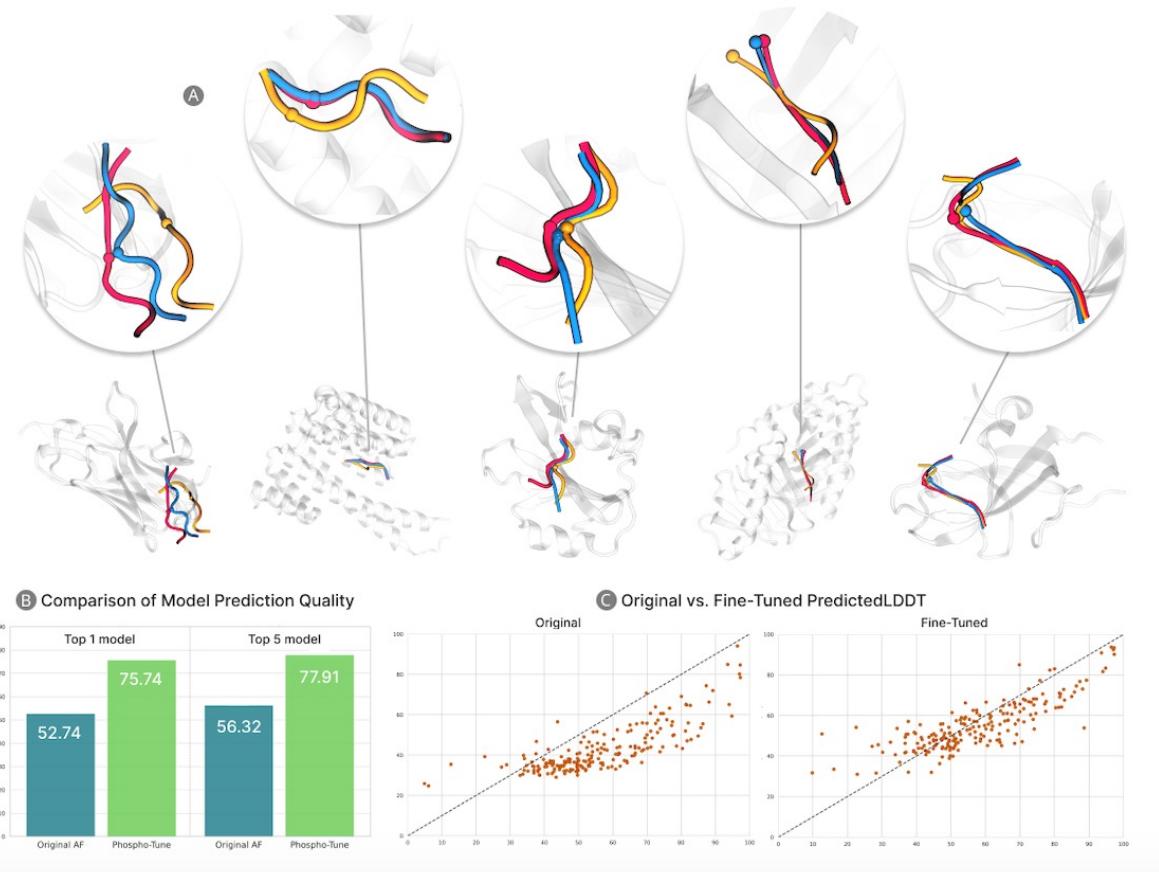
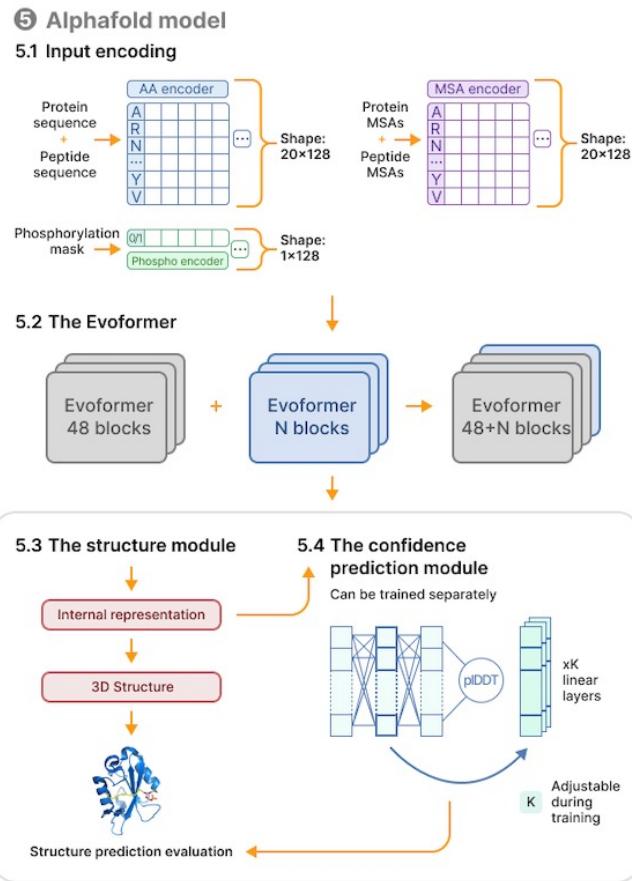
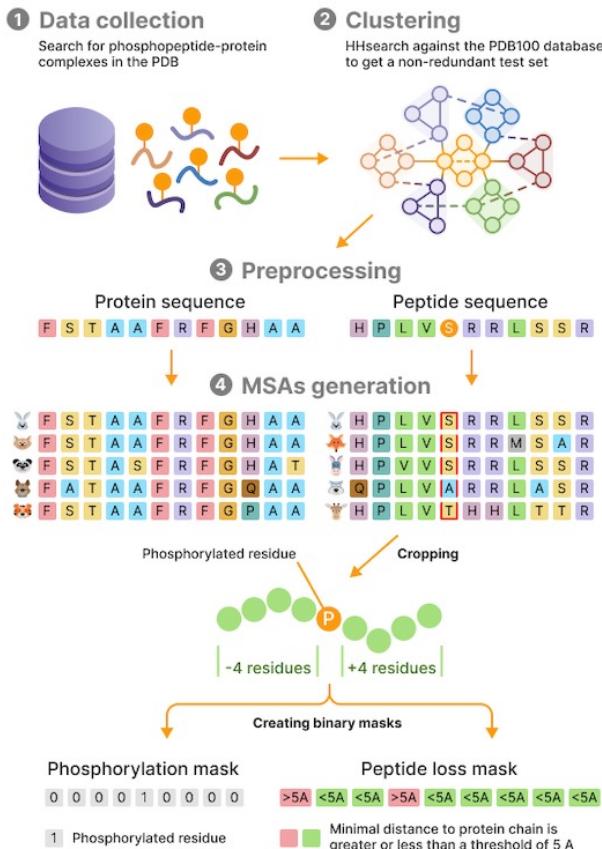


Currently in process of determining the X-ray structure of protein-ligand complex;

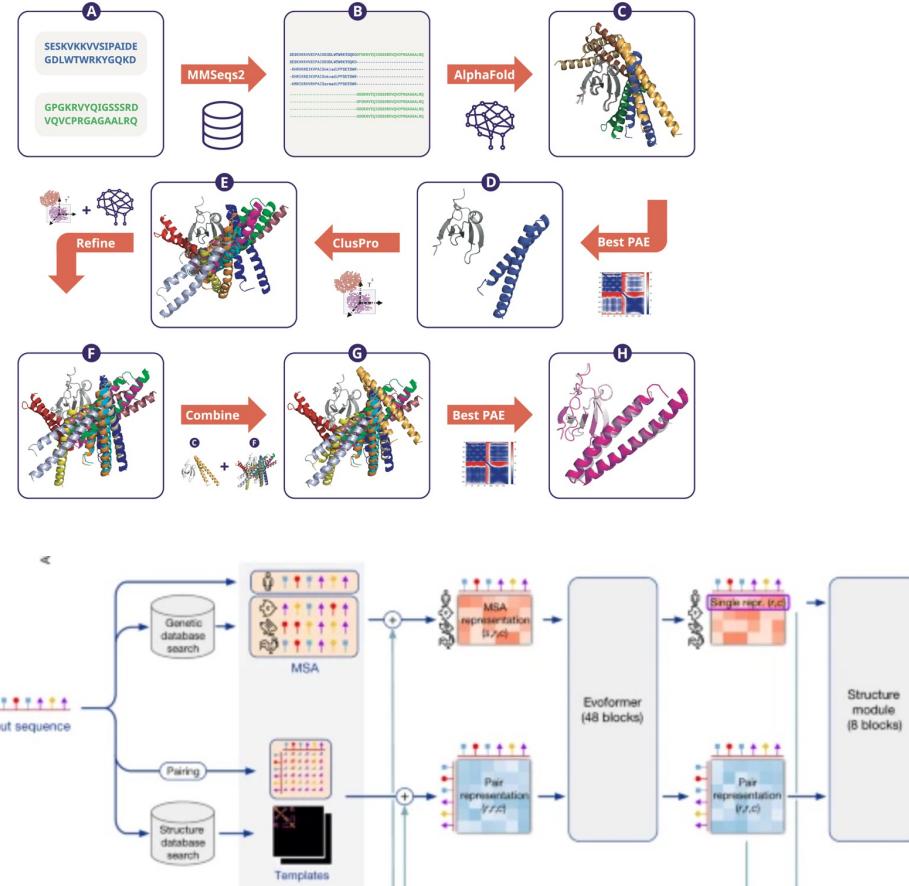
In collaboration with Professor Peter J. Tonge (Chemistry Dept.)  
And Qun Liu (Brookhaven National Laboratory)  
Alexander Tropsha (UNC Chapel Hill)



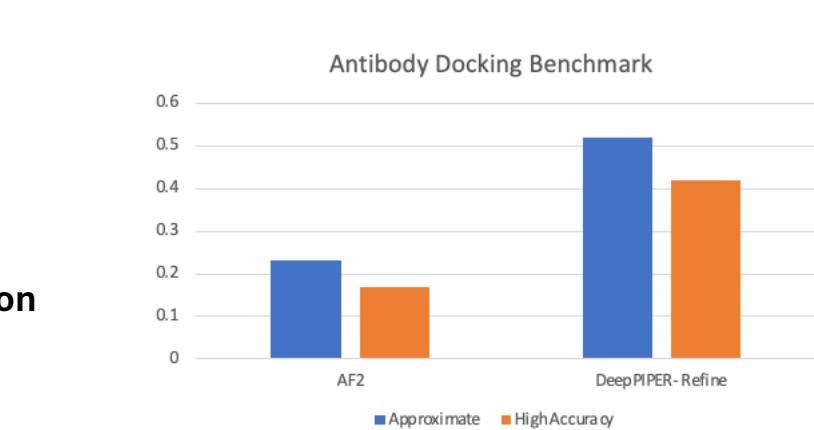
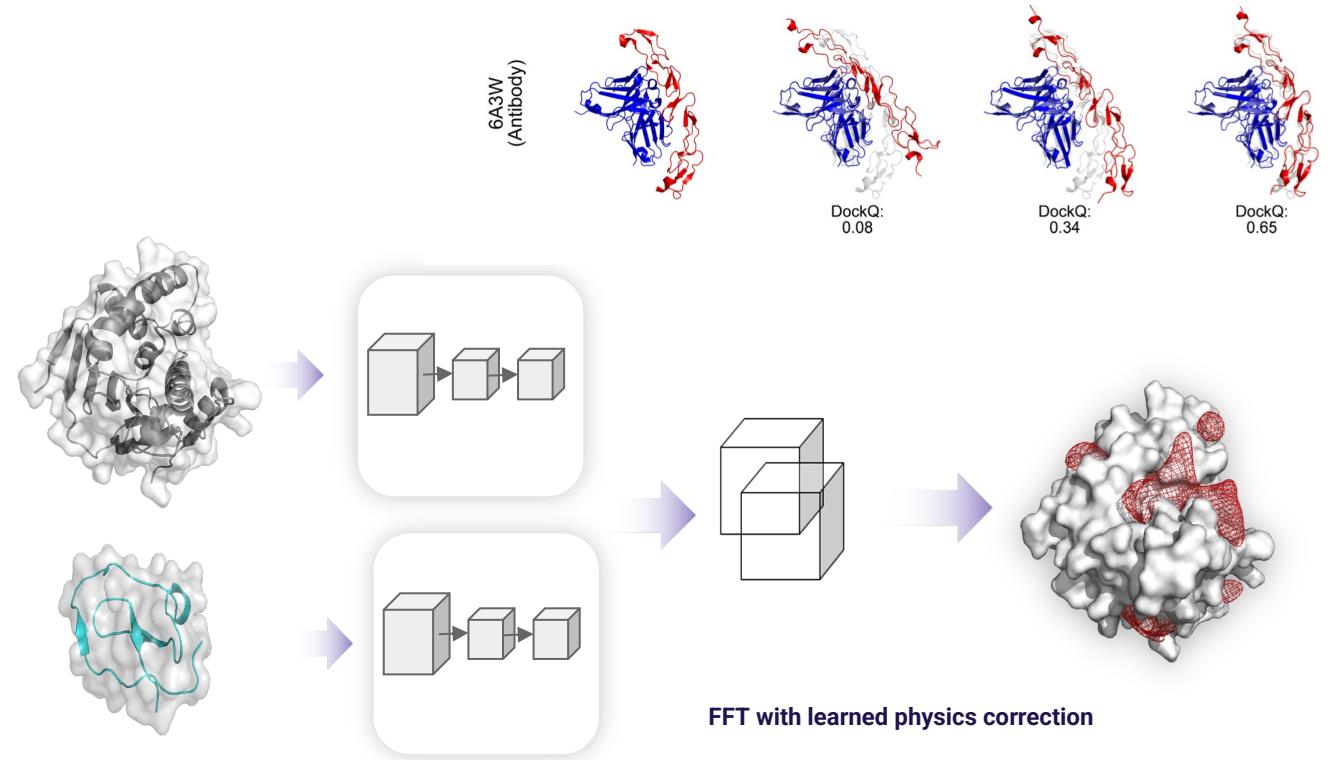
# Modeling Phosphorylated interactions



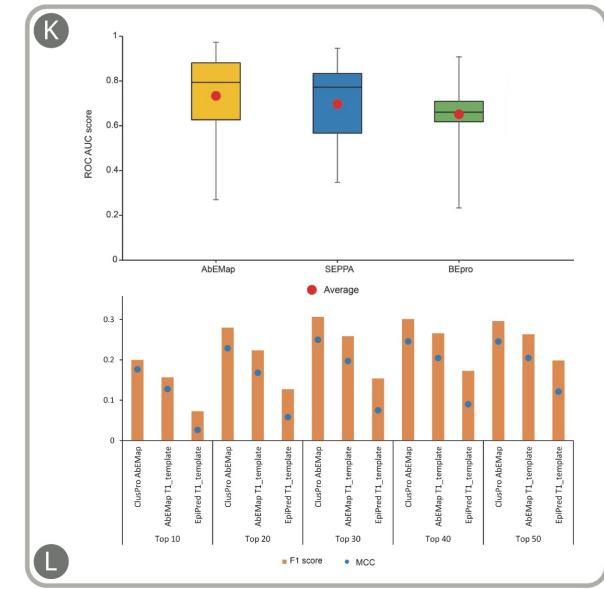
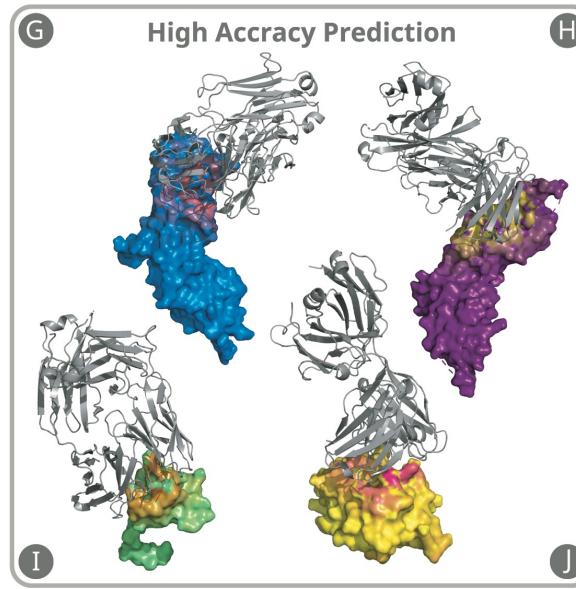
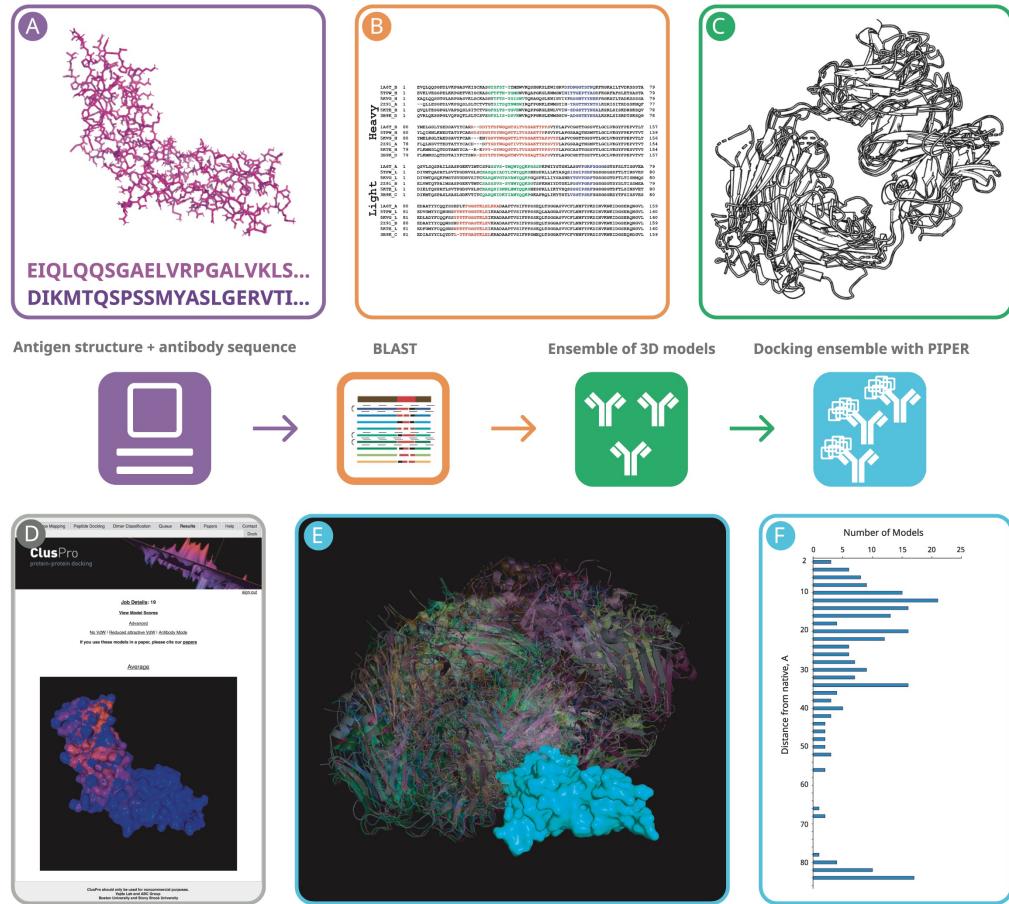
# Combination of FFT based architecture and AF for modeling Antigen Antibodies



**Top performers in CAPRI – worldwide blind protein docking competition**



# High Accuracy Epitope Detection



# ClusPro server 20000 users; FTMap server: over 5000 registered users

The screenshot shows the ClusPro web interface. At the top, there is a navigation bar with links for Dock, Queue, Results, API, Preferences, Downloads, Admin, Papers, Help, and Contact. Below the navigation bar is a large, colorful 3D surface plot representing a protein-protein docking result. A "Sign out" link is located in the top right corner of this plot area. The main content area is titled "Dock". It contains fields for "Job Name" (with a placeholder "RNA") and "Server" (set to "katana"). Below these, a note states "Accepted PDB Input: 20 standard amino acids and RNA (as receptor only), ref: RNA". There are two sections for "Receptor" and "Ligand", each with "PDB ID" input fields, "Upload PDB" buttons, and "Chains" input fields. A note below says "Whitespace separate desired chains. Leave chains blank to use all chains." There is also an "Advanced Options" link and a "Dock" button at the bottom. At the very bottom, a footer note reads: "ClusPro should only be used for noncommercial purposes. Structural Bioinformatics Lab Boston University".

The screenshot shows the FTMap web interface. The browser address bar shows "http://ftmap.bu.edu/". The main title is "FT-Map:A Small Molecule Mapping Server". The navigation bar includes Queue, Results, Preferences, Examples, Help, Papers, and Contact. The "Papers" tab is currently selected. The main content area features the "FTMap protein mapping" logo and a 3D visualization of a protein surface with several small molecules docked on it. A "Welcome to FTMAP Server!!" message is displayed. To the right, a detailed description of the service is provided: "Computational solvent mapping is a powerful tool to understand interactions between proteins and solvent molecules. It docks small organic molecules on a protein surface, finds favorable binding positions, clusters the conformations of all prediction, and ranks the clusters on the basis of their average free energy. The low energy clusters are grouped into consensus sites and the largest consensus sites are able to identify active or ligand binding sites. The docked fragments can also be served as the building blocks for fragment-based drug design." Below this text is a 3D molecular model showing binding sites labeled S4, S1, OX, and S1'.

# Modeling PROteolysis TArgeting Chimeras (PROTACs)

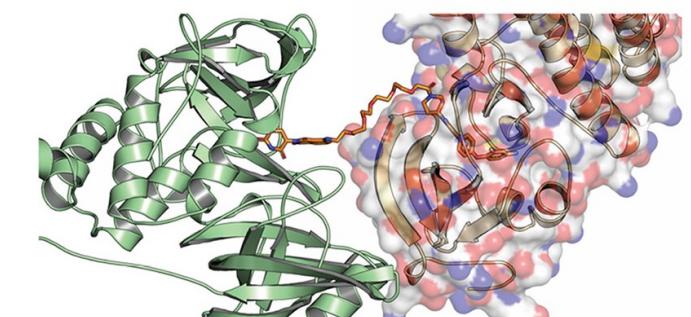
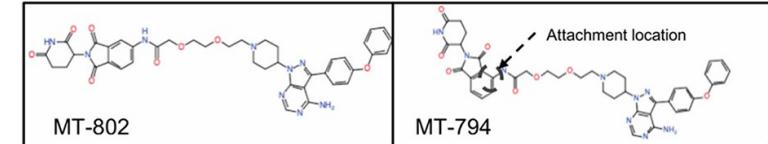
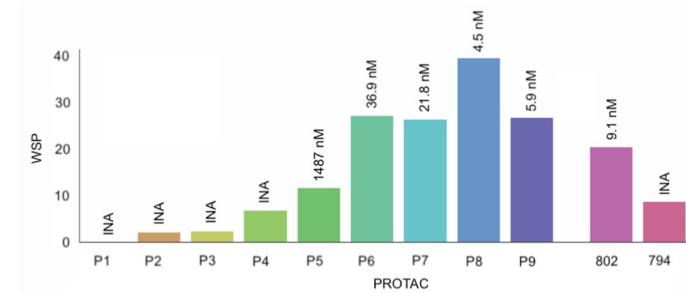
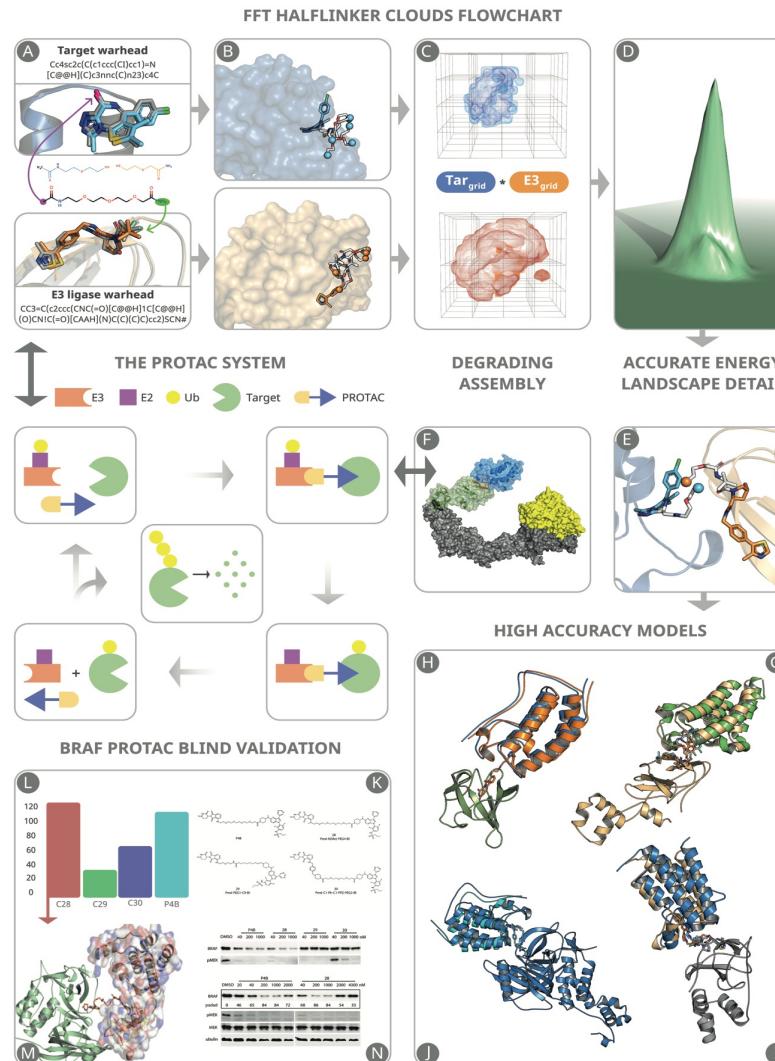
Goal: hijack ubiquitin-proteasome system to degrade target protein

We want to aid PROTAC design:

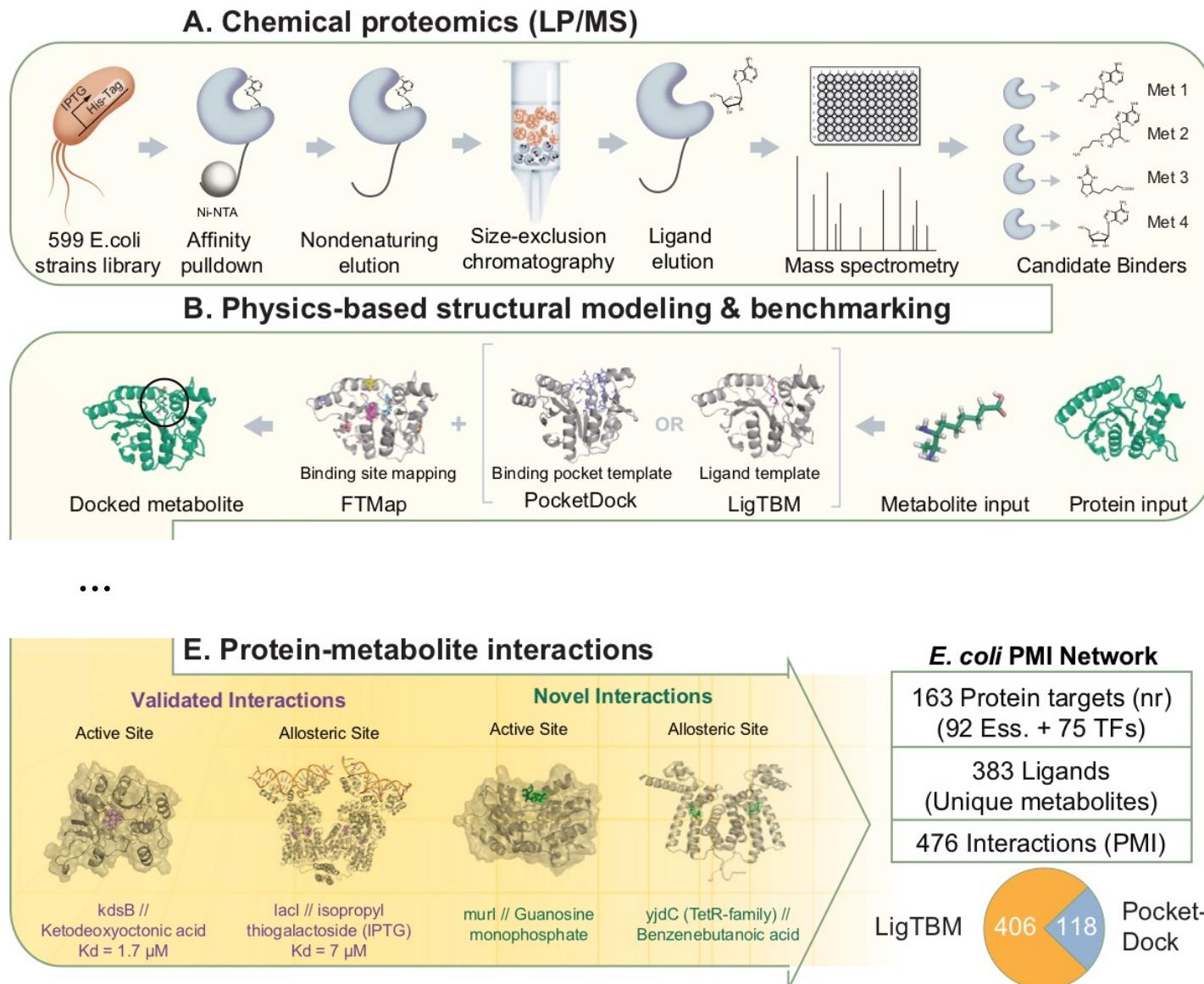
- PROTAC ternary complex structure
- PROTAC efficiency

Challenging sampling problem:

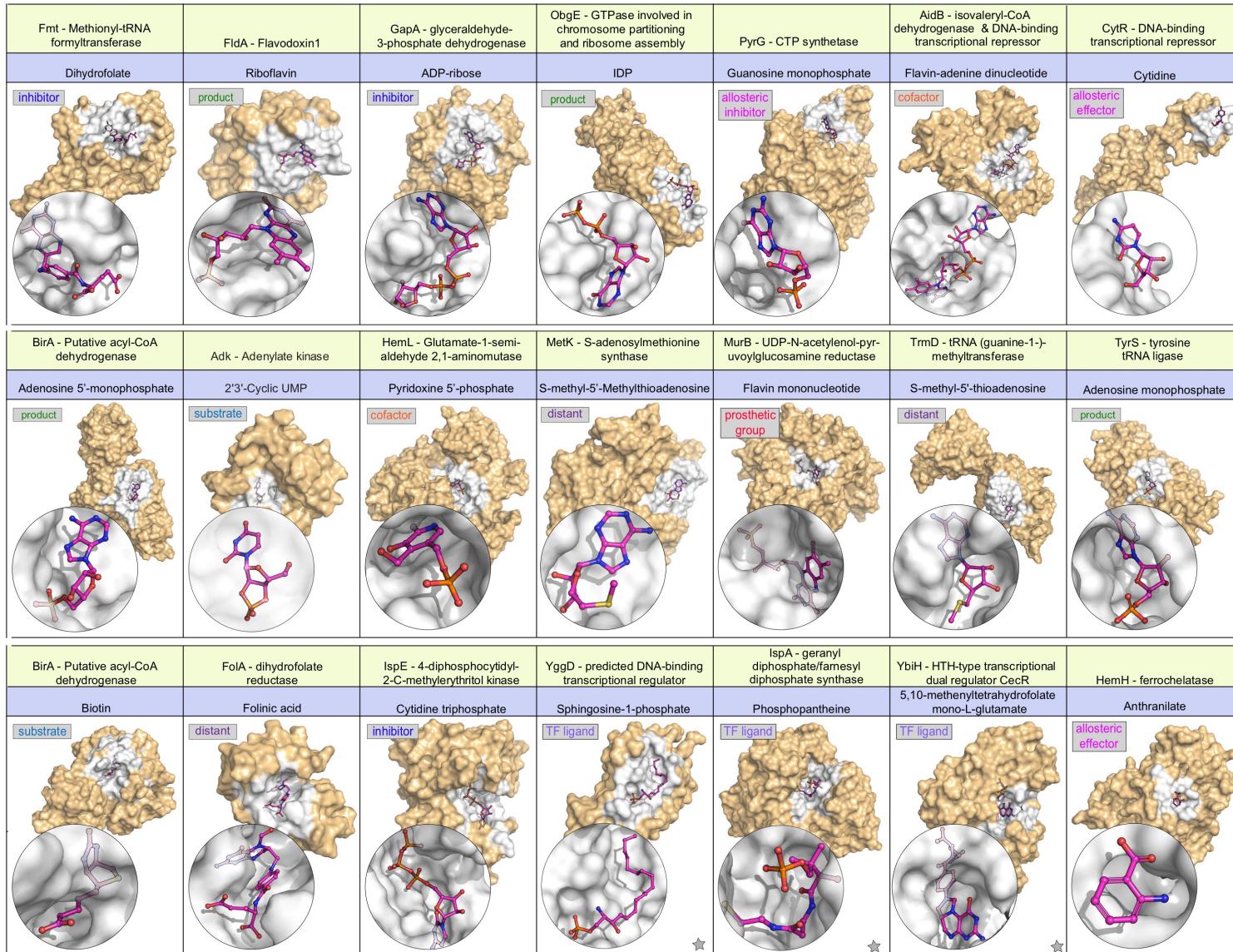
- PROTAC linker might have non-trivial chemistry and conformational space
- Multiscale modeling
- Non-native protein-protein interaction
- Suboptimal interface



# Large-scale mapping of native protein-metabolite interactions in E. coli using Mass Spec & LigTBM



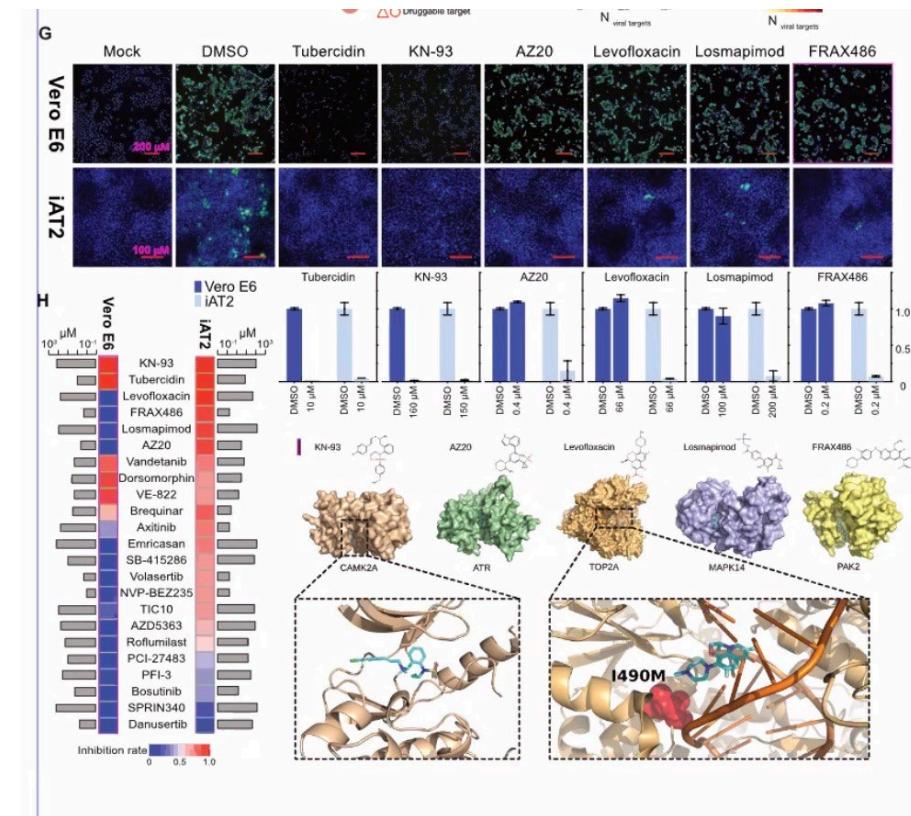
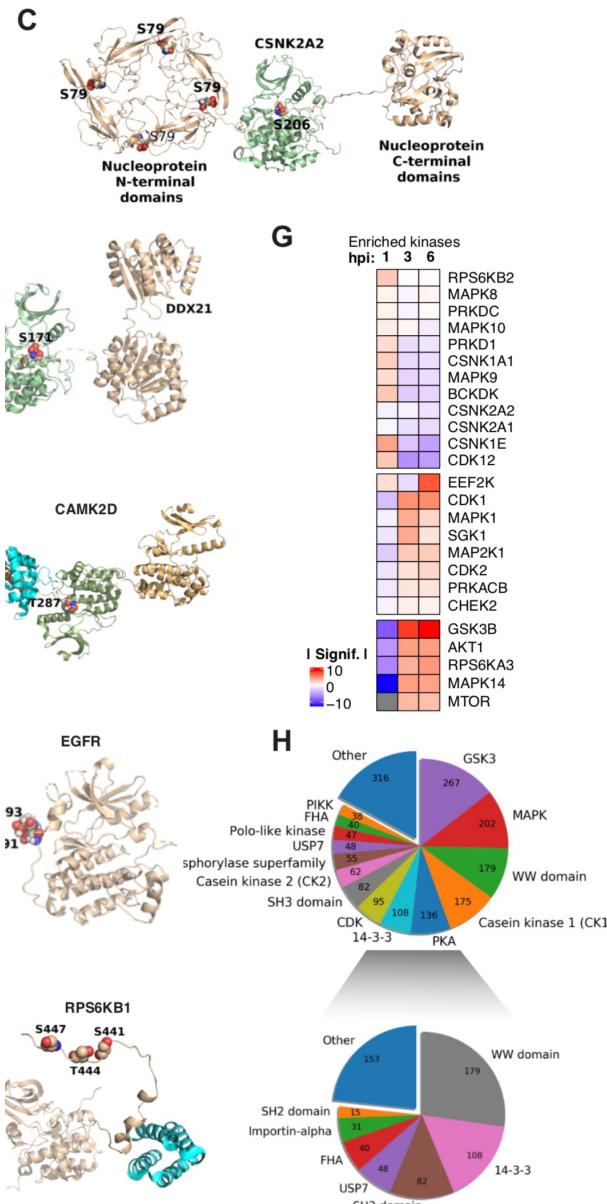
# Known and predicted protein-metabolite interactions & SPR validation



## Surface Plasmon Resonance assay

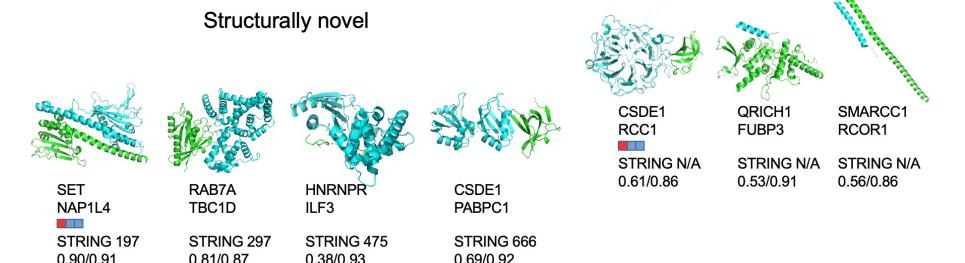
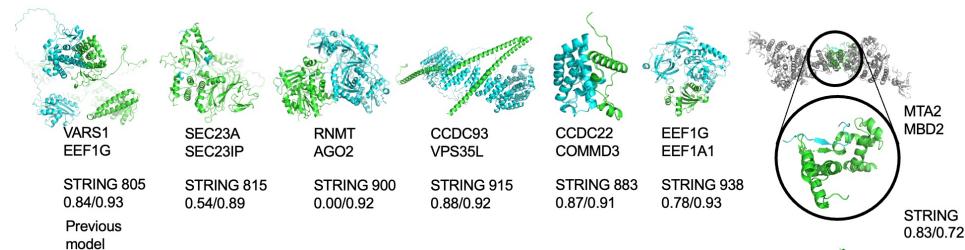
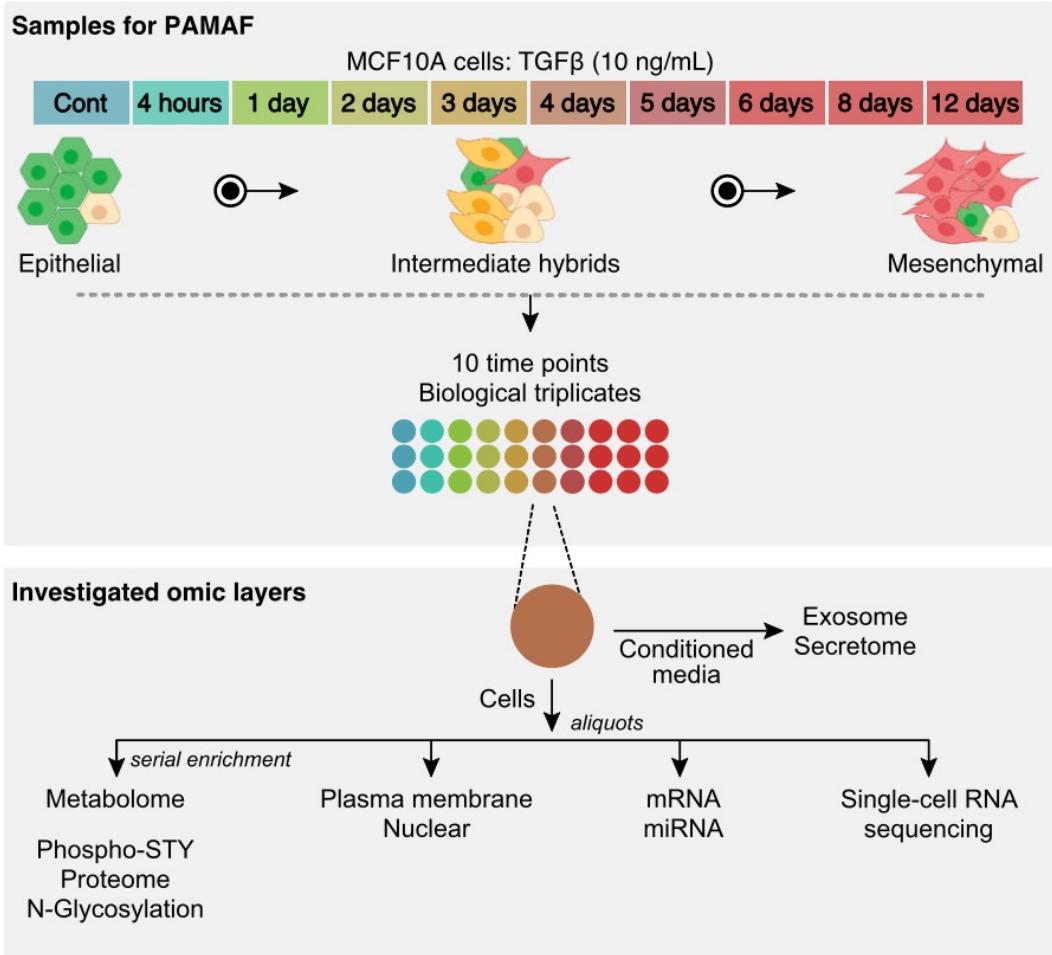
	KD (nM)
HemH - 4-aminobenzamide	483
YjdC - Oleic acid	3.8
PurB - Phenazine	61.6
FldA - [2,2'-Bipyridine]-5-carboxylic acid	192
IspA - (5-Fluoro-2-oxo-2,3-dihydro-1H-indol-3-yl)-acetic acid	54.5
ObgE - 1-isoquinolinyl(phenyl) methanol	119
FldA - Riboflavin	224
IspF - Phenazine	60.1
IspB - 1-(3,4-Dimethoxy-phenyl)-ethylamine	118
TrmD - S-methyl-5'-thioadenosine	14.9
MetK - S-methyl-5'-thioadenosine	0.355
UvrY - 4-chloro-2-hydroxybenzamide	170
PyrG - Guanosine	1250
IspE - Cytidine	67.7
MurB - Flavin mononucleotide	84500
HemH - Phenazine-Flavohemochromogen	6.241

# Phosphorylation effect of SARS-COV2 on infected lung cells – Target identification

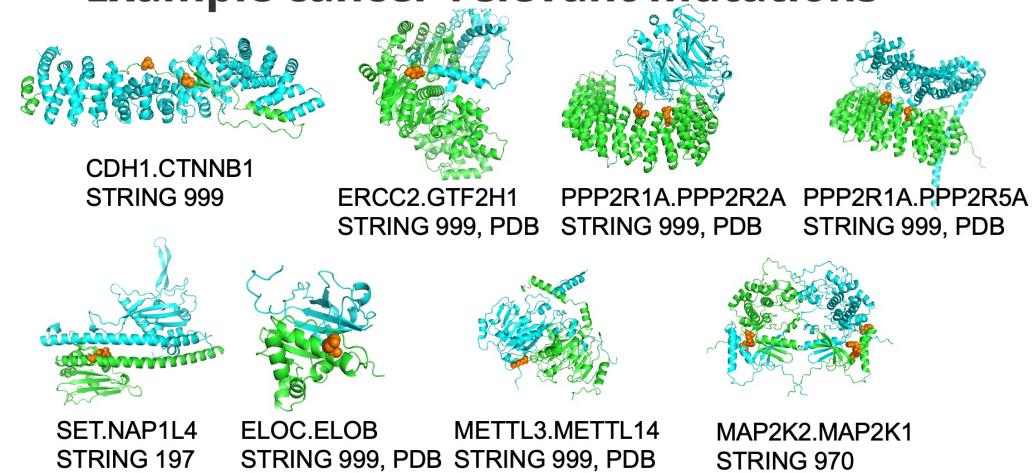


Hume et. al, Molecular Cell, 2021

# Understanding EMT

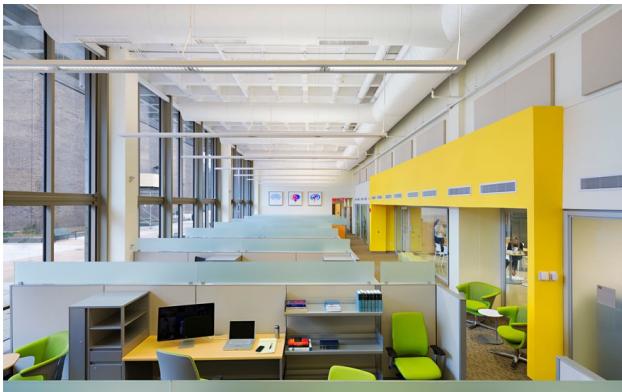


## Example cancer-relevant mutations



## ABC Lab members

Dr. Dzmitry Padhorny  
Sergei Kotelnikov  
Dr. Mikhail Ignatov  
Ernest Glukhov  
Ryota Ashizawa  
Xiaogang Li  
Dr. Mark Lukin  
George Jones  
Thu Nguyen  
Yimin Zhu  
Derara Haligeorgious  
Veranika Averkova  
Dmytro Kalitin



## SBU

Ken Dill  
Carlos Simmerling  
Ivet Bahar  
Peter Tonge  
David Thanassi  
Vageli Coutsias

## U of Toronto, OHSU

Andrew Emili

## BU

Sandor Vajda  
Adrian Whitty

## UNC

Alex Tropsha  
Tim Wilson

**NIH , NSF**  
**Computer time DOE INCITE leadership award**